



Tetrahedron Letters 47 (2006) 6259-6262

Tetrahedron Letters

## Daphnilongerine, an unprecedented fused pentacyclic ring system alkaloid from *Daphniphyllum longeracemosum* Rosenth.

Lin Li, a,b Hong-ping He, Ying-tong Di, Suo Gao and Xiao-jiang Hao a,\*

<sup>a</sup>State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, PR China <sup>b</sup>Graduate School of Chinese Academy of Sciences, Beijing 100039, PR China

Received 22 February 2006; revised 22 June 2006; accepted 26 June 2006

Abstract—An unusual yuzurine-type alkaloid daphnilongerine (1), with an unprecedented fused pentacyclic skeleton in addition to seven known ones, daphnigracine (2), daphnezomine R, daphnigraciline, yuzurine, longistylumphyllines A, daphnilongeranin C, and calycinine A, was isolated from the fruits of *Daphniphyllum longeracemosum*. The structure and relative stereochemistry of 1 were determined by spectroscopic analysis.

© 2006 Elsevier Ltd. All rights reserved.

Daphniphyllum alkaloids are a group of highly complex and diversified polycyclic alkaloids, which have been the challenging subjects of natural products, biogenetic pathways, and synthetic programs. Despite more than 100 Daphniphyllum alkaloids having been isolated from the genus Daphniphyllum by Kobayshi, Jossang, Yue, Bodo, and their co-workers, 1–5 there is still a great interest for the discovery of unique ring system alkaloids from a biogenetic point of view. 6,7

Daphniphyllum longeracemosum Rosenth. is an evergreen tree distributed in the Yunnan Province of China. Yue and co-workers have reported four new Daphniphyllum alkaloids daphnilongeranins A–D, along with four known ones, which were isolated from the leaves and stems of this plant. 4a Our investigation has shown that the major alkaloids of the fruits of this plant, however, are of the yuzurine type. 1 Herein we reported an unprecedented pentacyclic fused ring system alkaloid daphnilongerine (1), together with seven known alkaloids, daphnigracine (2), daphnezomine R, daphnigraciline, by yuzurine, blongistylumphyllines A, daphnilongeranin C, and calycinine A, which was isolated from the fruits of D. longeracemosum Rosenth.

 $\label{lem:condition} \textit{Keywords: Daphniphyllum longeracemosum; Daphniphyllum alkaloid; Daphnilongerine.}$ 

The fruits of *D. longeracemosum* were extracted with 95% EtOH. The crude extract was adjusted with 10% HCl to pH  $\sim$  2. The acidic mixture was defatted with petroleum ether (PE), and then extracted with CHCl<sub>3</sub>. The aqueous phase was basified to pH  $\sim$  10 with saturated Na<sub>2</sub>CO<sub>3</sub> and extracted with CHCl<sub>3</sub> to obtain crude alkaloids (8.0 g). The crude alkaloids were subjected to a silica gel column (CHCl<sub>3</sub>/MeOH, 1:0  $\rightarrow$  0:1, v/v) to give four fractions (1–4). Fraction 2 was chromatographed over a silica gel column (PE/EtOAc/Et<sub>2</sub>NH, 10:1:0.25 to 1:1:0.25) followed by silica gel (PE/acetone, 8:2) to yield daphnilongerine (1, 0.0004%).

Daphnilongerine (1) was isolated as an optically active colorless solid,  $[\alpha]_D^{21.5}$  +65.0 (c 0.98, CH<sub>3</sub>OH). The ESIMS showed the molecular ion [M+H]<sup>+</sup> at m/z 402 and the HRESIMS established the molecular formula  $C_{24}H_{36}NO_4$  (m/z 402.2641, [M+H]<sup>+</sup>; calcd: 402.2644). Thus, the structure of 1 possessed eight degrees of unsaturation. The IR absorption spectrum suggested the presence of hydroxyl (3443 cm<sup>-1</sup>) and two carbonyls (1735 cm<sup>-1</sup> and 1705 cm<sup>-1</sup>). The 1D and 2D NMR spectra (CD<sub>3</sub>OD, Table 1) displayed 24 carbon signals consisting of two carbonyls, two  $sp^2$  quaternary carbons assignable to one tetrasubstituted double bond, two  $sp^3$  quaternary carbons, six  $sp^3$  methines (two methines at  $\delta_C$  42.3 were observed by HSQC and HMBC), eight  $sp^3$  methylene, and four methyl groups. Among them, one methylene ( $\delta_C$  60.0;  $\delta_H$  2.62 and 2.08), one methine ( $\delta_C$  71.9;  $\delta_H$  2.87–2.86), and one methyl ( $\delta_C$  43.1;  $\delta_H$ 

<sup>\*</sup>Corresponding author. Tel.: +86 871 5223263; fax: +86 871 5219684; e-mail: haoxj@mail.kib.ac.cn

Table 1. <sup>1</sup>H and <sup>13</sup>C NMR, HMBC, and ROESY correlations of 1 in CD<sub>3</sub>OD

	$\delta_{\mathrm{H}}^{\mathrm{a}}$ , multi, $J$ (Hz)	${\delta_{ m C}}^{ m b}$	$\begin{array}{c} HMBC \\ H \rightarrow C \end{array}$	$\begin{array}{c} ROESY \\ H \rightarrow H \end{array}$
1α	2.62 (br d, 11.7)	60.0 (CH <sub>2</sub> )	5, 7, 8, 9	15, 1β
β	2.08 (br d, 11.7)		7, 8, 9, 13, 24	3, 1α, 13α
2		218.1 (C)		
3	3.25 (dd, 9.2, 5.4)	44.8 (CH)	2, 4, 6, 7	18, 24, 1 $\beta$ , 4 $\alpha$ , 19, 20
$4\alpha$	1.51 (dd, 9.2, 13.4)	34.1 (CH <sub>2</sub> )	2, 5, 6, 7, 8	$21\alpha$ , 3, $13\beta$ , $4\beta$
β	2.21–2.15 (m)		2, 3, 5, 21	$21\alpha$ , $21\beta$ , $4\alpha$
5		51.6 (C)		
6	2.04-2.00 (m)	42.3 (CH)	5, 7, 8, 11	21β, 7, 12β
7	2.87–2.86 (m)	71.9 (CH)	1, 2, 4, 5, 24	24, 6
8	. ,	47.6 (C)		
9		145.6 (C)		
10		138.9 (C)		
11α	2.24–2.21 (m)	26.4 (CH <sub>2</sub> )	9, 10	
β	1.93–1.89 (m)		17	12α
12α	2.29-2.27 (m)	25.9 (CH <sub>2</sub> )	5, 10	11β, 12β
β	1.74–1.71 (m)		5, 6, 11	12α, 6
13α	1.65 (dd, 14.0, 8.7)	39.8 (CH <sub>2</sub> )	1, 8, 9, 14, 15, 22	14, 13β, 1β
β	2.45 (dd, 14.0, 6.9)		1, 5, 8, 14, 22	$21\alpha$ , $13\alpha$ , $4\alpha$
14	2.97-2.91 (m)	42.3 (CH)	9, 15, 22	15, 13α
15	3.53-3.50 (m)	53.7 (CH)		14, $1\alpha$ , $16\alpha$
16α	1.95–1.92 (m)	29.3 (CH <sub>2</sub> )	9, 10, 15	15, 17α, 16β
β	1.32–1.28 (m)		14, 15	17β, 16α
17α	2.59-2.54 (m)	41.6 (CH <sub>2</sub> )		17β, 16α
β	2.35–2.31 (m)		9, 10, 15, 16	17α, 16β
18	2.86-2.80 (m)	40.9 (CH)	2, 19, 20	3, 19, 20
19	1.09 (d, 7.0)	19.4 (CH <sub>3</sub> )	2, 18, 20	18
20	1.06 (d, 7.0)	18.8 (CH <sub>3</sub> )	2, 18, 19	18
21α	3.85 (br d, 11.5)	65.7 (CH <sub>2</sub> )	4, 5, 6, 8	$21\beta$ , $13\beta$ , $4\beta$ , $4\alpha$
β	3.59 (br d, 11.5)	• •	4, 5, 6, 8	$21\alpha$ , $4\beta$ , $6$
22		177.5 (C)		•
23	3.62 (3H, s)	51.8 (CH <sub>3</sub> )	22	
24	2.27 (3H, s)	43.1 (CH <sub>3</sub> )	1, 7	3, 7

<sup>&</sup>lt;sup>a</sup> Measured at 400 MHz.

2.27) were ascribed to those linking to a nitrogen. The chemical shift of the methylene at  $\delta_C$  65.7 suggested its linking to an oxygen. Since the carbonyl groups and the only double bond accounted for 3 out of 8 unsaturations, 1 was inferred to possess five rings.

By an extensive comparison of <sup>1</sup>H and <sup>13</sup>C NMR data of **1** with daphnigracine, <sup>8</sup> **1** was suggested to possess a partial structure (Fig. 1, part A) of daphnigracine. However, the presence of an unprecedented fused pentacyclic ring system for **1** has not been observed from other *Daphniphyllum* alkaloids. So, 2D NMR experi-

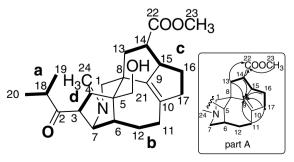


Figure 1. <sup>1</sup>H-<sup>1</sup>H COSY, TOCSY, and part A of 1.

ments were required to determine the whole structure of 1.

Analysis of the <sup>1</sup>H-<sup>1</sup>H COSY, TOCSY, HSQC, and HMBC spectra established four fragments, a (C-18 to C-19 and C-20), **b** (C-12 to C-6 and C-11,C-6 to C-7), c (C-14 to C-13 and C-15, C-15 to C-14 and C-16, C-16 to C-15 and C-17), and d (C-3 and C-4) drawn with bold bonds (Fig. 1). In part A, C-9 attaching to C-15 could be determined by the HMBC correlations of H-14/C-9, H-16α/C-9. The HMBC correlations of H- $11\alpha/C-10$ , C-9, H-11 $\beta/C-17$  and H-17 $\beta/C-10$ , C-9 suggested that C-10 connected fragments c and b. The correlations between C-22 ( $\delta_{\rm C}$  177.5) with the methoxyl ( $\delta_{\rm C}$ 51.8,  $\delta_{\rm H}$  3.62) and H-14 ( $\delta_{\rm H}$  2.97–2.91), indicated the only ester group was located at C-14 ( $\delta_{\rm C}$  42.3). H-6 ( $\delta_{\rm H}$  2.04–2.00) displayed correlations with C-7, C-5 and C-11 ( $\delta_{\rm C}$  26.4) in HMBC. The connectivity between C-1 and C-13 through C-8 was provided by HMBC correlations of  $H_2$ -1/C-8, H-1 $\beta$ /C-13,  $H_2$ -13/C-1. The linkage of two quaternary carbons C-8 and C-9 could be determined by the HMBC correlations between H-13 $\alpha$  and C-9, H<sub>2</sub>-1 and C-9. A ketone carbonyl  $(\delta_C 218.1)$  connected fragments **a** and **d** on the basis of its HMBC correlations with H-18 ( $\delta_{\rm H}$  2.86–2.80),  $H_3$ -19 ( $\delta_H$  1.09),  $H_3$ -20 ( $\delta_H$  1.06), H-3 ( $\delta_H$  3.25) and

<sup>&</sup>lt;sup>b</sup> Measured at 100 MHz.

H<sub>2</sub>-4 ( $\delta_{\rm H}$  2.21–2.15 and 1.51). HMBC correlations of H-3/C-7 ( $\delta_{\rm C}$  71.9), H-3/C-6 ( $\delta_{\rm C}$  42.3), H-7/C-4 ( $\delta_{\rm C}$  34.1), and H-4α/C-7 indicated the connectivity of C-3 to C-7. HMBC correlations of H<sub>3</sub>-24 to C-1 ( $\delta_{\rm C}$  60.0) and C-7, H-7 to C-1 and C-24, H-1 to C-24 and C-7 suggested that C-1, C-7, and C-24 were all linked to the same nitrogen atom. C-21 ( $\delta_{\rm C}$  65.7), C-4, C-8 ( $\delta_{\rm C}$  47.6) and C-6 were attached to the C-5 ( $\delta_{\rm C}$  51.6) on the grounds of HMBC correlations of H<sub>2</sub>-21/C-4, C-6, C-8, C-5, H<sub>2</sub>-4/C-5, H-6/C-8. The above data could be explained by the presence of a new carbon ring connecting C-3–C-4–C-5–C-6–C-7 in 1 in contrast to other yuzurine-type alkaloids. Thus, the gross planar structure of daphnilongerine was assigned as shown in Figures 1 and 2.

The relative stereochemistry of 1 was deduced by the ROESY spectrum as shown in the computer-generated three-dimensional drawing (Fig. 3). The ROESY correlations of H-13 $\alpha$ /H-14, H-14/H-15, H-15/H-16 $\alpha$ , and H-16 $\alpha$ /H-17 $\alpha$  indicated that H-14, H-15 were  $\alpha$ -oriented. H-6 and H-7 were assigned  $\beta$ -configuration on the basis of the correlation of H-6/H-12  $\beta$ , H-6/21 $\beta$ , H-6/H-7. The correlation of H-3/H-4 $\alpha$  determined H-3 was  $\alpha$ -oriented.

The configuration of daphnilongerine (1) may also be deduced by the inspection of the proposed biosynthesis. From a biogenetic point of view, it could be admitted

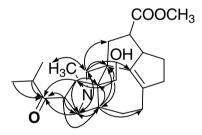
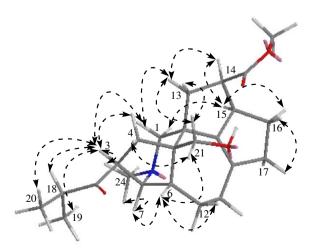


Figure 2. Key HMBC correlations of 1 ( $H \curvearrowright C$ ).



**Figure 3.** Key ROESY correlations and conformation generated from computer modeling (Chem. Draw. 9.0 3D).

Scheme 1. Biogenetic pathway proposed for daphnilongerine (1).

that yuzurine-type alkaloids represented by daphnilongerine (1) and daphnigracine (2)<sup>8</sup> may be produced from an intermediate A derived from squalene. The several-step oxidative reaction may convert A to imine B, the latter underwent plausible Mannich reaction to give daphnilongerine (1). On the other hand, the condensation reaction of A yields hemiketal product daphnigracine (2) (Scheme 1).

A cytotoxicity assay showed that compound 1 was not active against the human glioblastoma (U251), human lung cancer (A549), the acute myelogenous leukemia (HL60), human liver carcinoma (BEL-7402), murine leukemia (P388), and murine melanoma (B16) cell lines.

## Acknowledgements

We are grateful to Professor Xun Gong, Kunming Institute of Botany, Chinese Academy of Sciences (CAS), for the collection and identification of the plant material.

## Supplementary data

1D and 2D NMR; HRESIMS; and IR. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2006.06.144.

## References and notes

- For reviews of *Daphniphyllum* alkaloids, see: (a) Yamamura, S.; Hirata, Y. In *The Alkaloids*; Manske, R. H. F., Ed.; Academic Press: New York, 1975; Vol. 15, p 41; (b) Yamamura, S. In *The Alkaloids*; Brossi, A., Ed.; Academic Press: New York, 1986; Vol. 29, p 265; (c) Kobayashi, J.; Morita, H. In *The Alkaloids*; Cordell, G. A., Ed.; Academic Press: New York, 2003; Vol. 60, p 165.
- (a) Morita, H.; Ishioka, N.; Takatsu, H.; Shinzato, T.; Obara, Y.; Nakahata, N.; Kobayashi, J. Org. Lett. 2005, 7, 459–462; (b) Takatsu, H.; Morita, H.; Ya-Ching, S.; Kobayashi, J. Tetrahedron 2004, 60, 6279–6284; (c) Morita, H.; Takatsu, H.; Ya-Ching, S.; Kobayashi, J. Tetrahedron Lett. 2004, 45, 901–904; (d) Morita, H.; Kobayashi, J. Org. Lett. 2003, 5, 2895–2898; (e) Kobayashi, J.; Takatsu, H.;

- Shen, Y. C.; Morita, H. Org. Lett. 2003, 5, 1733–1736; (f) Morita, H.; Takatsu, H.; Kobayashi, J. Tetrahedron 2003, 59, 3575–3579; (g) Kobayashi, J.; Ueno, S.; Morita, H. J. Org. Chem. 2002, 67, 6546–6549; (h) Morita, H.; Yoshida, N.; Kobayashi, J. J. Org. Chem. 2002, 67, 2278–2282; (i) Morita, H.; Kobayashi, J. Tetrahedron 2002, 58, 6637–6641; (j) Kobayashi, J.; Inaba, Y.; Shiro, M.; Yoshida, N.; Morita, H. J. Am. Chem. Soc. 2001, 123, 11402–11408; (k) Morita, H.; Yoshida, N.; Kobayashi, J. J. Org. Chem. 2000, 65, 3558–3562; (l) Morita, H.; Yoshida, N.; Kobayashi, J. Tetrahedron 1999, 55, 12549–12556.
- 3. Jossang, A.; Bitar, H. E.; Pham, V. C.; Sévenet, T. *J. Org. Chem.* **2003**, *68*, 300–304.
- (a) Yang, S. P.; Zhang, H.; Zhang, C. R.; Cheng, H. D.; Yue, J. M. J. Nat. Prod. 2006, 69, 79–82; (b) Zhan, Z. J.; Zhang, C. R.; Yue, J. M. Tetrahedron 2005, 61, 11038–11045; (c) Chen, X.; Zhan, Z. J.; Yue, J. M. Helv. Chim. Acta 2005, 88, 854–860; (d) Zhan, Z. J.; Yang, S. P.; Yue, J. M. J. Org. Chem. 2004, 69, 1726–1729; (e) Yang, S. P.; Yue, J. M. Org. Lett. 2004, 6, 1401–1404; (f) Yang, S. P.; Yue, J. M. J. Org. Chem. 2003, 68, 7961–7966.

- (a) Bitar, H. E.; Nguyen, V. H.; Gramain, A.; Sévenet, T.; Bodo, B. *Tetrahedron Lett.* 2004, 45, 515–518; (b) Bitar, H. E.; Nguyen, V. H.; Gramain, A.; Sévenet, T.; Bodo, B. *J. Nat. Prod.* 2004, 67, 1094–1099.
- (a) Wallace, G. A.; Heathcock, C. H. J. Org. Chem. 2001, 66, 450-454; (b) Heathcock, C. H. Proc. Natl. Acad. Sci. USA 1996, 93, 14323-14327; (c) Heathcock, C. H.; Joe, D. J. Org. Chem. 1995, 60, 1131-1142; (d) Heathcock, C. H.; Kath, J. C.; Ruggeri, R. B. J. Org. Chem. 1995, 60, 1120-1130; (e) Heathcock, C. H. Angew. Chem. 1992, 104, 675-691; (f) Heathcock, C. H. Angew. Chem., Int. Ed. Engl. 1992, 31, 665-681.
- Ruggeri, R. B.; Hansen, M. M.; Heathcock, C. H. J. Am. Chem. Soc. 1988, 110, 8734–8736.
- (a) Yamamura, S.; Lamberton, J. A.; Irikawa, H.; Okumura, Y.; Hirata, Y. *Chem. Lett.* 1975, 9, 923–926; (b) Yamamura, S.; Lamberton, J. A.; Irikawa, H.; Okumura, Y.; Toda, M.; Hirata, Y. *Bull. Chem. Soc. Jpn.* 1977, 50, 1836–1840.
- Hao, X. J.; Zhou, J.; Node, M.; Fuji, K. Yunnan Zhiwu Yanjiu, 1993, 15, 205–207.